

INTERACTION STUDIES WITH DNA

II. THE BINDING OF "DIMERIC" AND "POLYMERIC" ROSANILINE BY SODIUM THYMONUCLEATE AND THE METACHROMATIC EFFECT

by

P. D. LAWLEY

*Chester Beatty Research Institute, Institute of Cancer Research,
Royal Cancer Hospital, London (England)*

The changes in absorption spectrum of metachromatic basic dyestuffs in aqueous solution and in presence of negatively charged colloids, and their dependence on the ratio of concentration of dyestuff to that of colloid, have been well defined from the qualitative point of view¹, particularly in the case of the interaction between toluidine blue and nucleic acids^{2, 3, 4}.

Rosaniline hydrochloride is a basic dye which shows no deviation from Beer's Law⁵ in aqueous solution at least up to a concentration of about 10^{-4} M, whereas in presence of *ca.* one equivalent of sodium thymonucleate a strong metachromasy is exhibited^{6, 7}. This absence of dimerization or polymerisation in aqueous solution simplifies the problem of a more complete analysis of the changes in spectrum associated with metachromasy, and the purpose of the present communication is to suggest such analysis and to report experimental evidence in support of it.

EXPERIMENTAL

Sodium deoxyribonucleate (DNA) was a preparation from calf thymus by the method of BUTLER CONWAY AND JAMES⁸ (P = 7.25 %, H₂O = 19.9 %). Rosaniline hydrochloride (E. Gurr & Co.) was twice recrystallized from water and dried.

Spectrophotometric measurements were made using the Uvispek instrument, with concentrations of rosaniline 1.1, 2.2 and $4.4 \cdot 10^{-5}$ M in $\frac{1}{2}$ and 1 cm cell thickness.

Stock solutions of DNA were 0.2 % in $5 \cdot 10^{-3}$ NaCl, and mixtures were made by adding the appropriate volumes of $2.2 \cdot 10^{-4}$ M rosaniline to the diluted DNA solutions.

Results reported were, unless otherwise stated, made with unthermostatted solutions (room temperature 23° C); the effect of raising the temperature of the solutions to 45° C, using a water-heated cell-housing, was not great although always in the direction of decreasing metachromasy *e.g.* with $2.2 \cdot 10^{-5}$ M rosaniline and $2.2 \cdot 10^{-5}$ M DNA, ϵ at 555 m μ was raised from $35.5 \cdot 10^3$ at 23° C to $38 \cdot 10^3$ at 45° C.

RESULTS

Fig. 1 shows typical absorption spectra due to rosaniline and their dependence on DNA concentration. Curve 1 is that due to rosaniline hydrochloride in water, (concentration denoted by R), with R = 1.1, 2.2 or $4.4 \cdot 10^{-5}$ M. The effect of large

References p. 332.

excess of DNA ($2.42 \cdot 10^{-3} M$) is shown by curve 2, while curves 3 and 4 show the meta-chromatic spectra induced by a concentration of DNA approximately equal to that of R (for curve 3, $R = 2.2 \cdot 10^{-5} M$, $DNA = 2.42 \cdot 10^{-5} M$; for curve 4, $R = 4.4 \cdot 10^{-5} M$, $DNA = 4.84 \cdot 10^{-5} M$). Curves 5 and 6 are values of the extinction coefficients for bound "dimeric" and "polymeric" rosaniline derived in the manner to be discussed below.

Figs. 2 and 3 show the variations in ϵ for rosaniline ($2.2 \cdot 10^{-5} M$) due to various concentrations of DNA; (abscissae denote the molar fraction $R/(R + DNA)$).

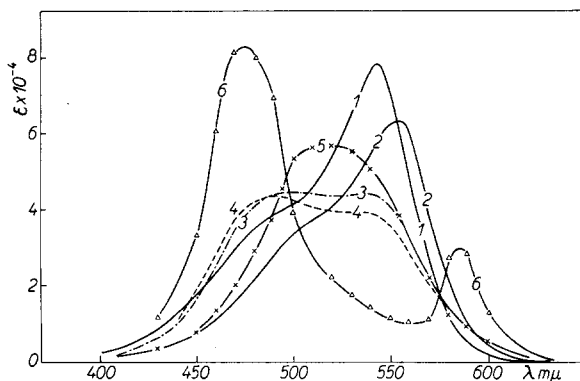


Fig. 1. Absorption spectra of rosaniline hydrochloride: (1) in water, (2) monomeric rosaniline bound to DNA (3) and (4) metachromatic spectra, (5) bound "dimer", (6) bound "polymer".

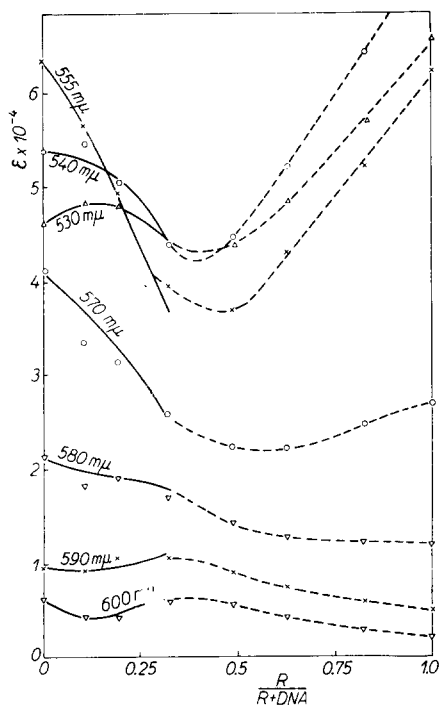


Fig. 2

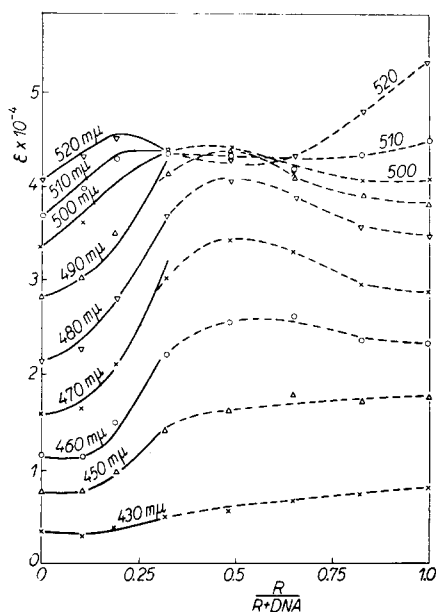


Fig. 3

Figs. 2 and 3. Variation of extinction coefficient due to rosaniline with the ratio of concentrations (rosaniline/rosaniline + DNA).

DISCUSSION

If the distance between the relevant binding sites (positions of negative charge) of a polymer is small (of the order of 3.5 Å), the occupation of neighbouring sites by dye-stuff cations, if sterically possible, as for those of planar configuration⁵ with their mole-

cular planes approximately at right angles to the polymer surface, will constitute the binding of "dimeric" or "polymeric" species, according as two or more adjacent sites are occupied. As MICHAELIS⁹ has pointed out, the configuration of DNA (as the fibrous Na salt) according to the model proposed by ASTBURY¹⁰ (or as modified by CRICK AND WATSON¹¹) is such as to present to the solution a surface of high charge density, with an array of primary phosphate groups separated by distances along the polymer chain of *ca.* 3.5 Å. MICHAELIS AND GRANICK¹² found that sodium nucleate, in contrast to most negative colloids, gave no metachromasy, or at most a weak effect, when the concentrations of nucleate and dyestuff were equal. However, this effect was due in the main to the lowering of the binding constant for the dyestuff by the presence of a large excess of Na⁺ ions⁷, as acetate buffer. In absence of added Na⁺, a strong metachromasy can be obtained with the usual dyestuffs, but this is displaced by adding metallic cations, as BANK AND BUNGENBERG DE JONG showed², divalent cations being some thirty times as effective as monovalent. These effects have been confirmed^{3,4}.

The origin of the metachromatic effect induced by DNA, (and similar considerations would be expected to apply to soap micelles, where metachromasy is exhibited when the concentration of micellar soap is approximately equal to that of the dyestuff, *i.e.* with small concentration of dye near the critical micelle concentration¹³, provided that this is greater than the dye concentration), would appear therefore to be somewhat as follows. For a given value of the degree of binding (β) (where β = the proportion of binding sites, assumed to be equal to the number of phosphate groups, occupied by rosaniline) the bound dye cations will be divisible into three groups, in the proportion $\beta_1:\beta_2:\beta_3$ respectively: (1) monomers, with sites on each side either unoccupied or occupied by Na⁺ ions; (2) "dimers", with one neighbouring site occupied by a dye cation; and (3) "polymers", with both neighbouring sites occupied by dye cations. The processes of "dimerization" or "polymerization" here denote juxtaposition of dye cations sufficiently close for Van der Waals' interaction¹⁴, such as to cause changes in the absorption spectrum of the dye.

Making the assumption that the binding sites are equivalent we obtain from simple statistical considerations that:

$$\beta_1 = \beta (1 - \beta)^2; \quad \beta_2 = 2\beta^2 (1 - \beta); \quad \text{and} \quad \beta_3 = \beta^3.$$

Further, $\beta = \beta_1 + \beta_2 + \beta_3$, may be related to the concentrations of free dye, (R_F), and of Na⁺ by the equation⁷:

$$\frac{(1 - \beta)}{\beta} (R_F) = \frac{1}{k'_R} = \frac{1}{k_R} + \frac{k_{Na}}{k_R} (Na^+).$$

The relation between the observed changes in absorption spectrum (observed extinction coefficient = ϵ) and the ratio of concentrations (R/DNA) is then obtained by introducing the extinction coefficient of free dye (ϵ°), of bound monomer (ϵ_1), bound "dimer" (ϵ_2), and bound "polymer" (ϵ_3):

$$\epsilon = \epsilon^\circ \left(1 - \frac{\beta \cdot \text{DNA}}{R}\right) + \epsilon_1 \beta_1 \left(\frac{\text{DNA}}{R}\right) + \epsilon_2 \beta_2 \left(\frac{\text{DNA}}{R}\right) + \epsilon_3 \beta_3 \left(\frac{\text{DNA}}{R}\right).$$

Of these quantities, apart from those known directly, ϵ_1 can be determined as the value of ϵ when (R/DNA) approaches zero; and for low ratios (R/DNA), in practice between 0 and 0.4, the binding constant k'_R in absence of added Na⁺ (apart from that derived by dilution of the stock DNA) may be assumed large enough for the approximation that all the R is bound, *i.e.* $\beta = (R/\text{DNA})$, to hold.

It becomes therefore possible to derive values of ϵ_2 and ϵ_3 on these assumptions, and these are shown in Fig. 1 as curves 5 and 6. Using these values, the calculated values of ϵ at various wavelengths have been inserted in Figs. 2 and 3, as the full curves for values of (R/DNA) less than 0.4; tolerable agreement with experimental data is obtained. The dotted curves for values of (R/DNA) greater than 0.4 show that the values of β become progressively less than the possible maximum, and that the binding constant decreases as (R/DNA) increases. However, the effect of decreasing concentration of DNA is in this case (with no added Na^+ apart from that of the stock solution, *i.e. ca.* twice the concentration of DNA) to lower the concentration of Na^+ to a level at which "denaturation" of DNA can occur. This process is shown by the increase in extinction coefficient of the absorption spectrum of DNA¹⁵, and results in a lowering of the affinity for binding of rosaniline⁷.

The forms of the absorption spectra of bound "dimeric" and "polymeric" dye are qualitatively as expected. The occurrence of a subsidiary peak, towards the longer-wavelength end of the spectrum, derived for the "polymer", corresponds with that observed during the salting-out of similar dyes¹⁶ and for crystalline hydrocarbons¹⁷, and shown to occur as part of the metachromatic spectrum by MICHAELIS AND GRANICK¹². In general, the present analysis may be said to confirm the view of MICHAELIS¹, that metachromasy results from the "dimerization" and "polymerization" of the basic dyes giving rise to absorption bands in the regions indicated.

ACKNOWLEDGEMENTS

The author's thanks are due to Dr. E. M. F. ROE and to Professors D. O. JORDAN, J. A. V. BUTLER and ALEXANDER HADDOW for their interest in this and related work, and to Professor BUTLER for the supply of DNA.

This work was supported by grants to the Chester Beatty Research Institute (Institute of Cancer Research, Royal Cancer Hospital) from the British Empire Cancer Campaign, Jane Coffin Childs Memorial Fund for Medical Research, the Anna Fuller Fund, and the National Cancer Institute of the National Institutes of Health, U.S. Public Health Service.

SUMMARY

1. The absorption spectra of rosaniline in aqueous solutions of sodium thymonucleate over the complete range of ratio of concentrations is reported.
2. The changes in spectrum of the dye are related to the proportions of dye bound according to a simple statistical analysis.
3. Absorption spectra for "dimeric" and "polymeric" bound rosaniline are deduced as an analysis of the metachromatic effect.

RÉSUMÉ

1. Les spectres d'absorption de la rosaniline dans des solutions aqueuses de thymonucléate de sodium dans tout le domaine du rapport des concentrations sont décrits.
2. Les modifications du spectre du colorant sont reliées aux proportions de colorant lié selon une analyse statistique simple.
3. Les spectres d'absorption de la rosaniline "dimérique" et "polymérique" liée sont déduits d'une analyse de l'effet métachromatique.

References p. 332.

ZUSAMMENFASSUNG

1. Das Absorptionsspektrum von Rosanilin in wässrigen Natriumthymonukleatlösungen im gesamten Bereich der Konzentrationsverhältnisse wird beschrieben.

2. Auf Grund einer einfachen statistischen Analyse werden die spektralen Veränderungen des Farbstoffes mit den Proportionen des gebundenen Farbstoffes in Verbindung gebracht.

3. Auf Grund der Analyse des metachromatischen Effektes werden Absorptionsspektren für "dimerisch" und "polymerisch" gebundenes Rosanilin gefolgert.

REFERENCES

- ¹ L. MICHAELIS, *J. Phys. and Colloid Chem.*, 54 (1950) 1.
- ² O. BANK AND H. G. BUNGENBERG DE JONG, *Protoplasma*, 32 (1939) 489.
- ³ N. WEISSMANN, W. H. CARNES, P. S. RUBIN AND J. FISHER, *J. Am. Chem. Soc.*, 74 (1952) 1423.
- ⁴ R. H. GARNER, D. O. JORDAN AND P. D. LAWLEY, *Ann. Rept. British Empire Cancer Campaign*, 29 (1951) 273.
- ⁵ S. E. SHEPPARD AND A. L. GEDDES, *J. Am. Chem. Soc.*, 66 (1944) 1995.
- ⁶ P. D. LAWLEY, *Thesis*, Nottingham, (1953).
- ⁷ P. D. LAWLEY, *Biochim. Biophys. Acta*, 19 (1956) 160.
- ⁸ J. A. V. BUTLER, B. E. CONWAY AND D. W. F. JAMES, *Trans. Faraday Soc.*, 50 (1954) 612.
- ⁹ L. MICHAELIS, *Cold Spring Harbor Symposia Quant. Biol.*, 12 (1947) 131.
- ¹⁰ W. T. ASTBURY, *Symposia Soc. Exptl. Biol.*, 1 (1947) 66.
- ¹¹ F. H. C. CRICK AND J. D. WATSON, *Nature*, 171 (1953) 737.
- ¹² L. MICHAELIS AND S. GRANICK, *J. Am. Chem. Soc.*, 67 (1945) 1212.
- ¹³ M. L. CORRIN, H. B. KLEVEN AND W. D. HARKINS, *J. Chem. Phys.*, 14 (1946) 480.
- ¹⁴ E. RABINOWITCH AND L. F. EPSTEIN, *J. Am. Chem. Soc.*, 63 (1941) 69.
- ¹⁵ R. THOMAS, *Biochim. Biophys. Acta*, 14 (1954) 231.
- ¹⁶ E. E. JELLEY, *Nature*, 138 (1936) 1009.
- ¹⁷ G. SCHEIBE, L. KANDLER AND H. ECKER, *Naturwiss.*, 26 (1938) 412.

Received June 23rd, 1955